



Mission pour les initiatives transverses et
interdisciplinaires (MITI)



Journée thématique

Fonctionnalisation de sondes et analyses de données

Sofiane EL-KIRAT-CHATEL



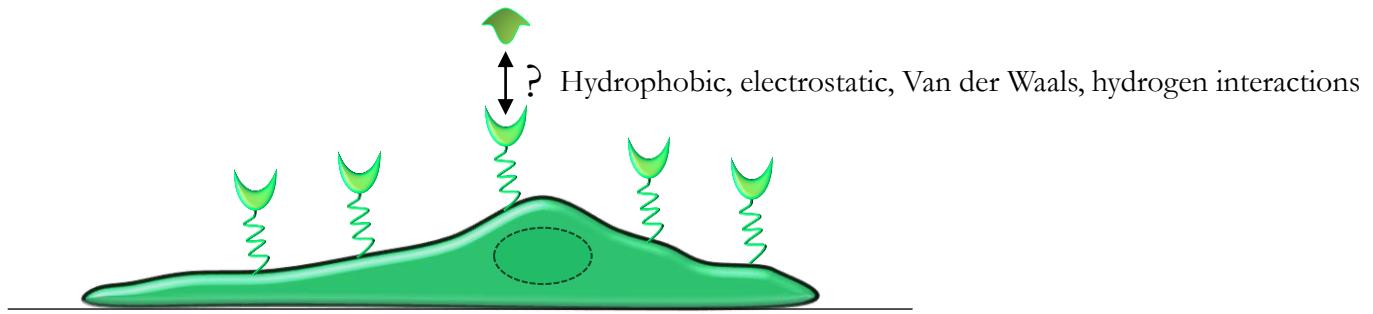
Chimie et Biologie des Membranes
et des Nano-objets
CBMN UMR 5248

Université
de BORDEAUX

Toulouse, 29 novembre 2023

Why measuring bio-interface interactions?

The bio-interface is the extreme limit between the intracellular compartment and the surrounding environment and governs signal transfer, molecular transport, and adhesion.



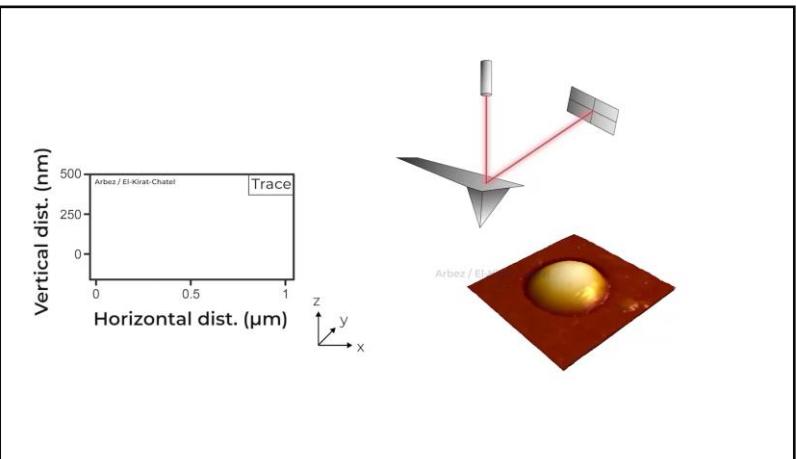
Probing ligand-receptor interactions and surface chemical properties to:

- Map the distribution of molecules / chemical properties
- Analyze molecular dynamics
- Determine biophysical properties/mechanics of molecules

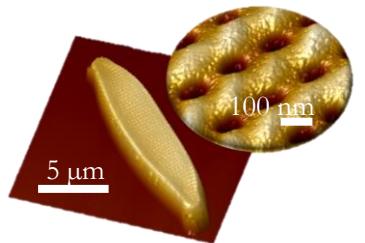
How to measure interaction forces?

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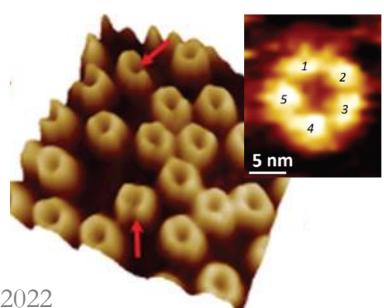
AFM: More than an imaging tool → Force spectroscopy to measure interaction forces



Nitzschia palea

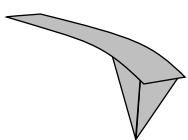


Prx decamer



Laviale *et al.* ACS AMI 2019

Beaussart *et al.* Nanoscale Horiz 2022

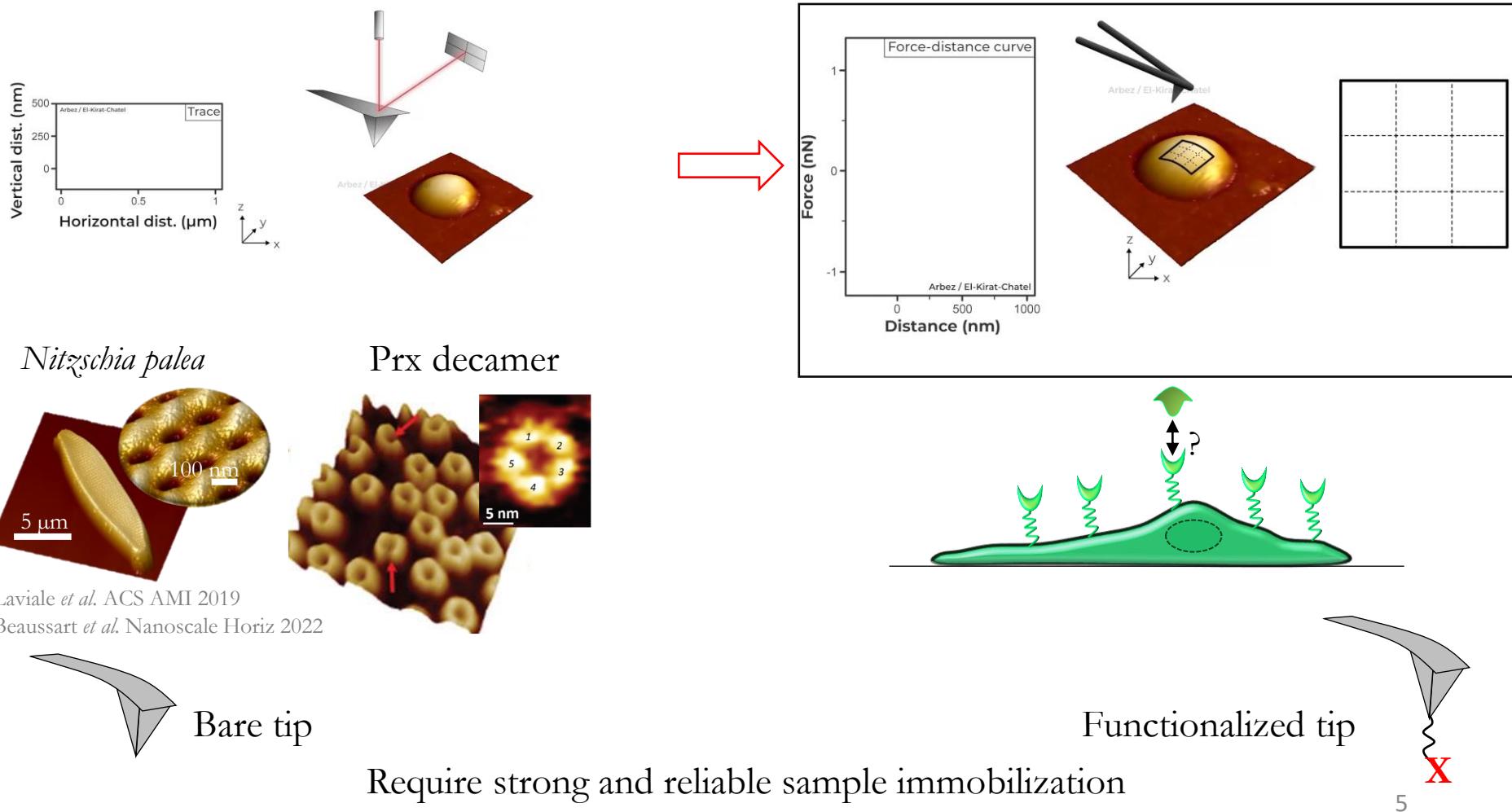


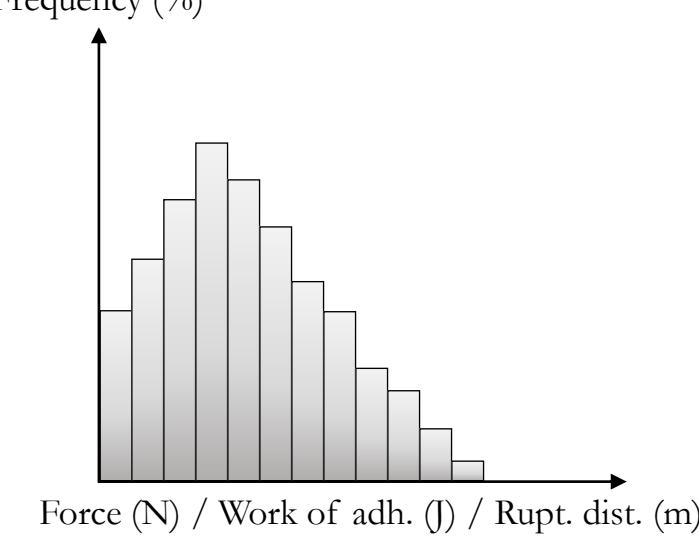
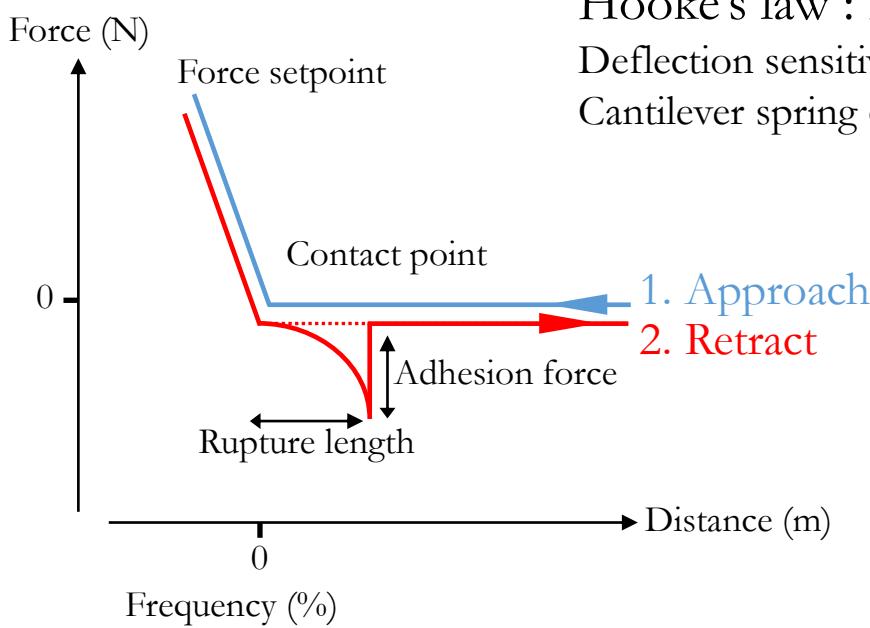
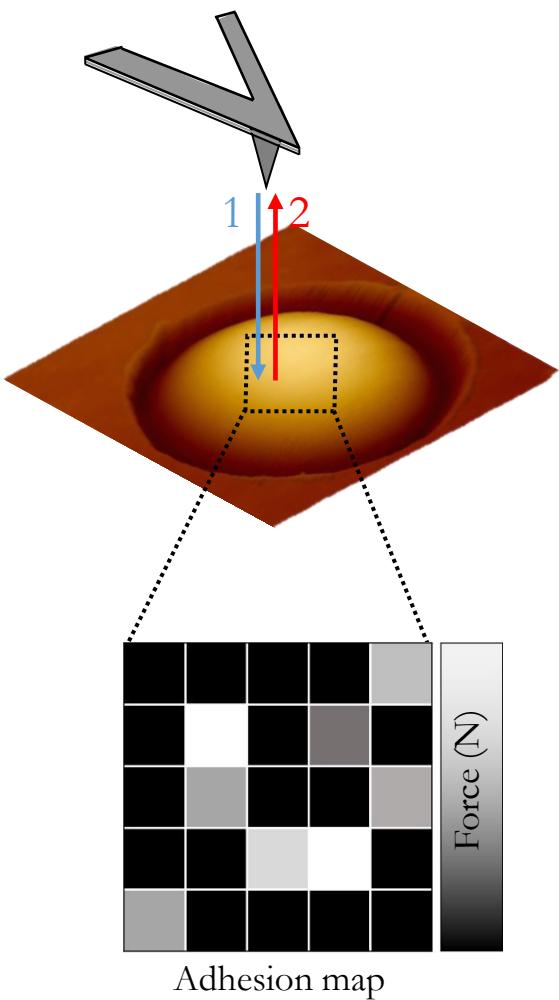
Bare tip

Require strong and reliable sample immobilization

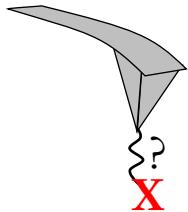
How to measure interaction forces?

AFM: More than an imaging tool → Force spectroscopy to measure interaction forces

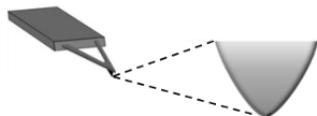
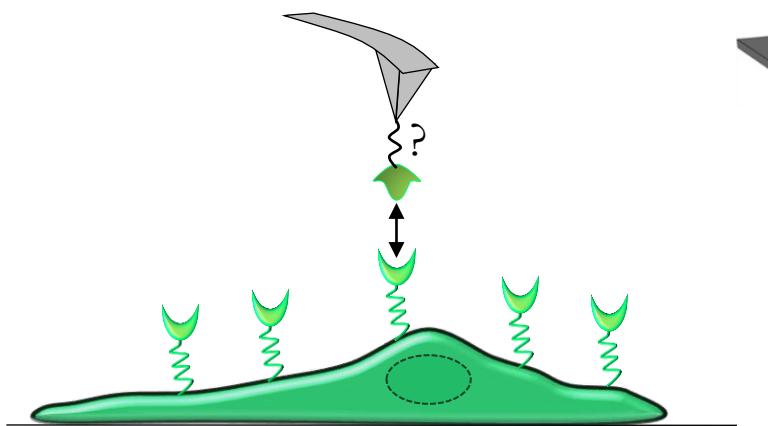




Hooke's law : $F = -k \times d$
Deflection sensitivity (nm/V)
Cantilever spring constant k (N/m)



How to functionalize AFM tips?



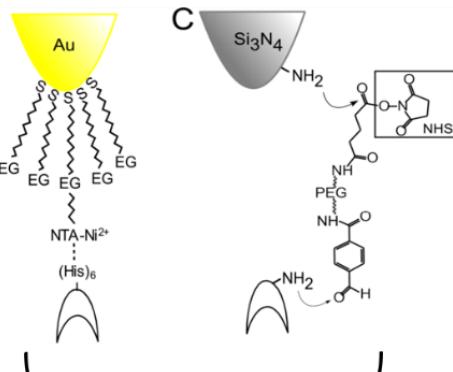
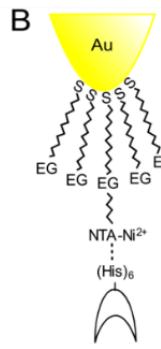
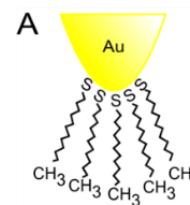
Tip functionalization examples

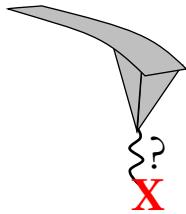
Nanomechanical
properties

Indentation
(courbe approche)

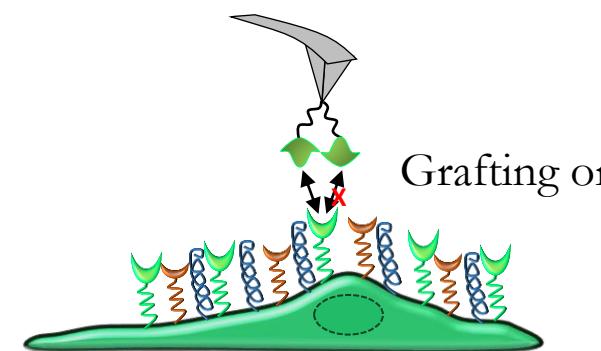
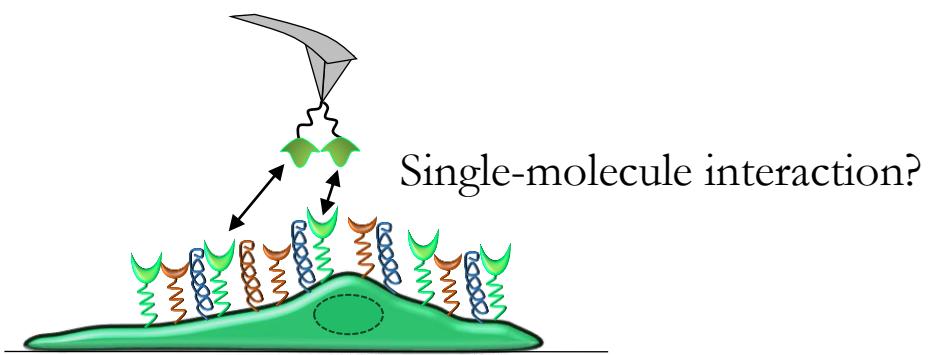
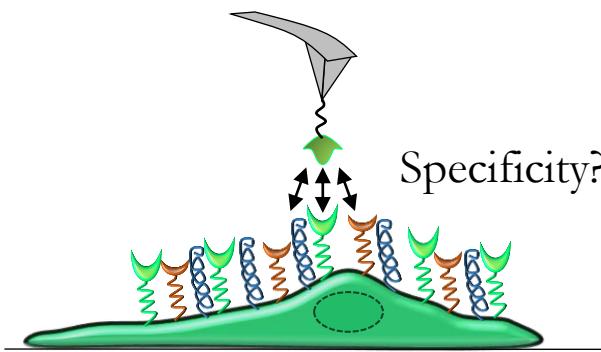
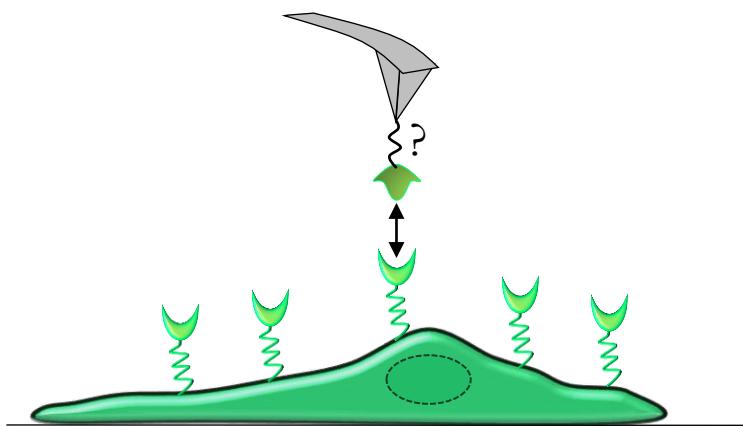
Chemical Force
Microscopy

Extension / interaction
(courbe retrait)





How to functionalize AFM tips?

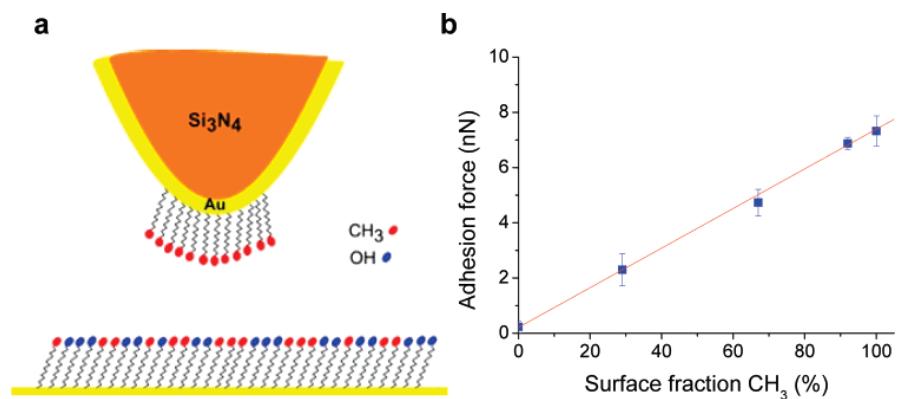
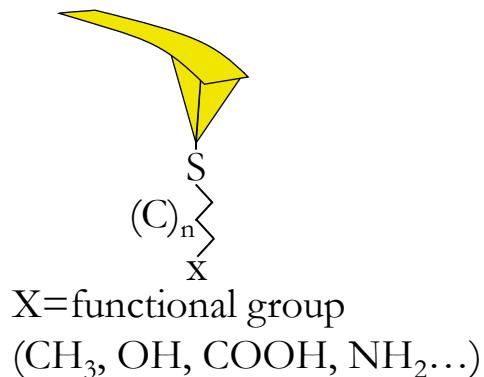


Tip functionalization protocol should ensure:

- Strong and flexible attachment
- Low grafting density for single-molecule
- No unspecific adhesion
- Preserved conformation and properties of the grafted object

Probing chemical properties of surfaces

Gold coated tip to graft
alkanethiol-terminated molecules



Dague et al. Nano letters 2007

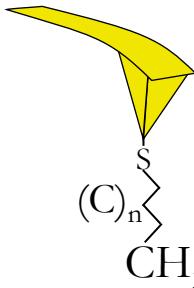
Protocol steps:

- AFM tip cleaning (UV/O_3 ; piranha...)
- Rinsing with ethanol + N_2 drying
- Immersion 4-12h in 1 mM solution of thiols in ethanol
- Rinsing with ethanol + N_2 drying

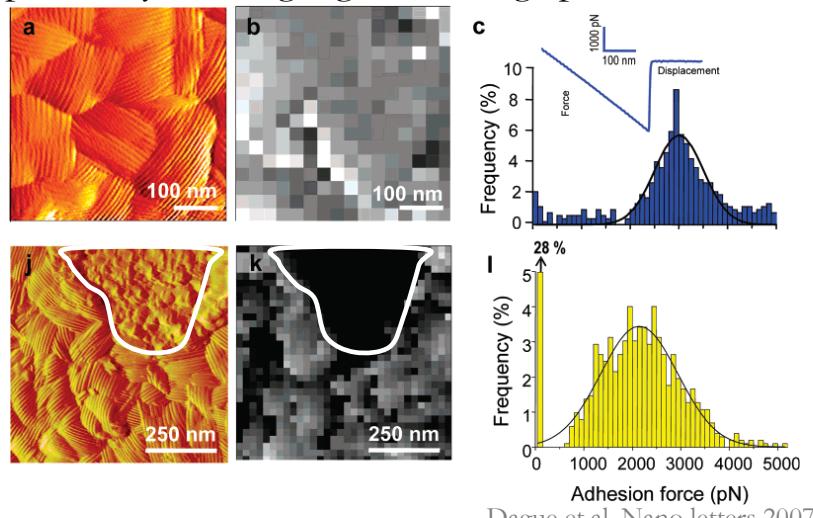
Example of thiols from Sigma:

- CH_3 : 1-dodecanethiol
- OH : 11-mercaptop-1-undecanol
- COOH : 16-mercaptophexadecanoic acid
- NH_2 : 1-Amino-1-undecanethiol

➤ Surface hydrophobicity of fungal germinating spores

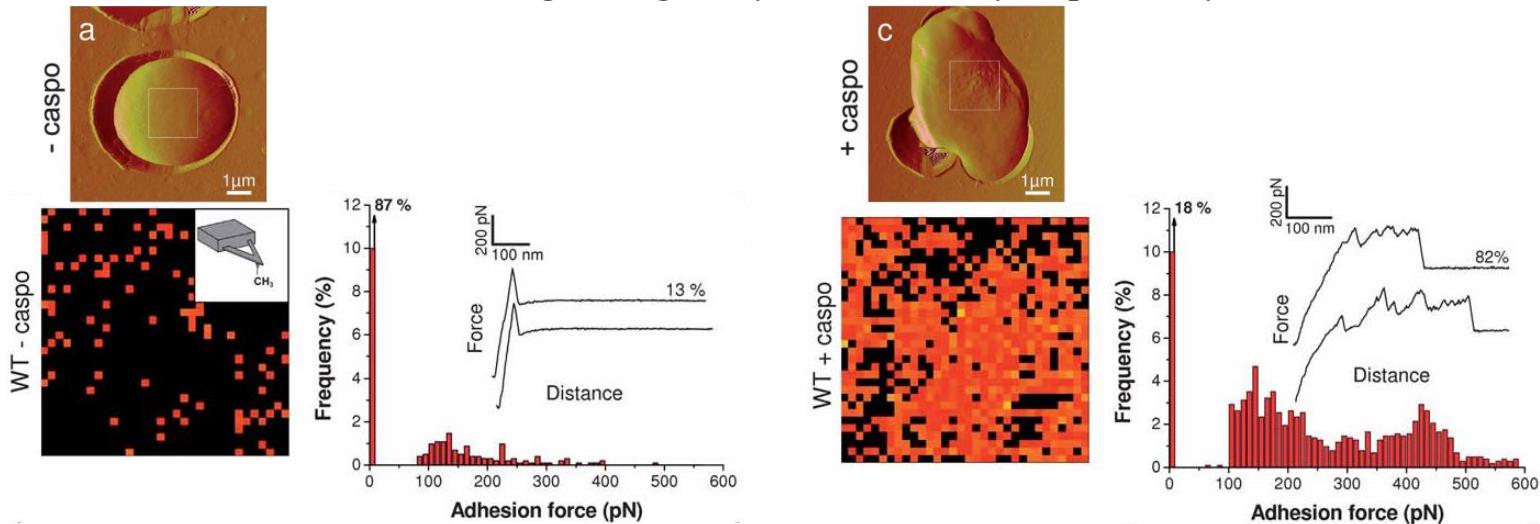


Hydrophobic tips coated
with 1-dodecanethiol



Dague et al. Nano letters 2007

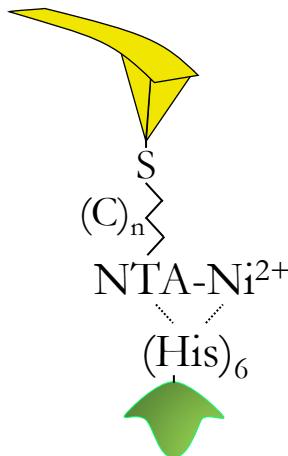
➤ Effects of antifungal drugs on yeast surface hydrophobicity



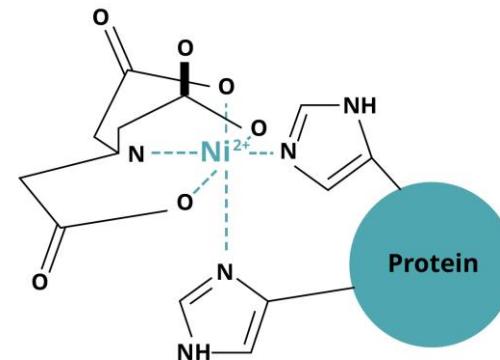
El-Kirat-Chatel et al. Nanoscale 2012

Limit: No molecular specificity

Gold coated tip to graft thiol-terminated molecules



Detection or oriented grafting of His-tagged proteins

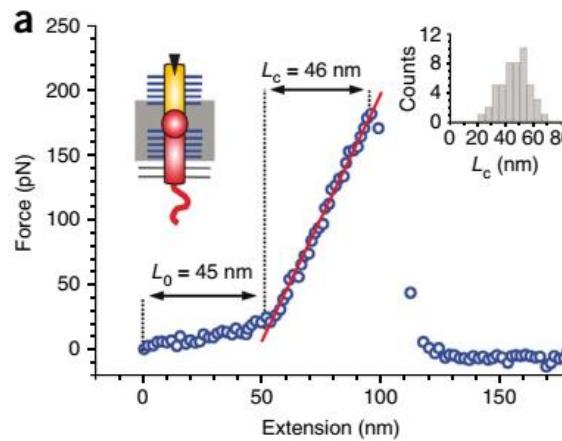
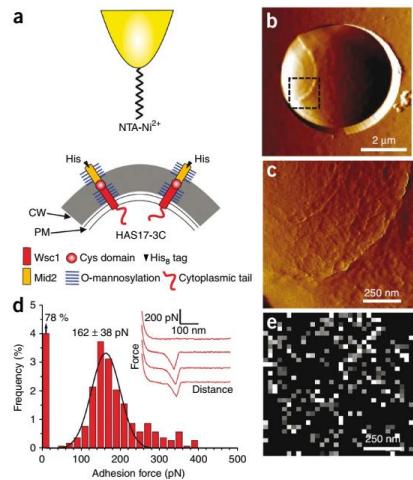


Protocol steps:

- AFM tip cleaning (UV/O₃; piranha...)
- Rinsing with ethanol + N₂ drying
- Immersion 4-12h in 0.01 mM NTA-terminated alkanethiols (or NTA-PEG alkanethiols) in ethanol (+spacers?)
- Rinsing with ethanol + N₂ drying
- Immerse 30 min in 80 mM NiSO₄
- Rinse gently with pure water (avoid drying)
For protein oriented grafting
- Add 100-200µL of His-tagged proteins (200µg/mL) 1-2h before gentle rinsing in buffer

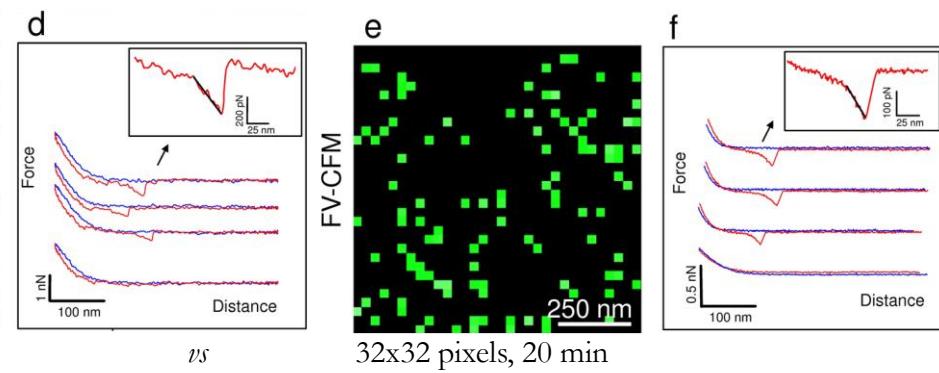
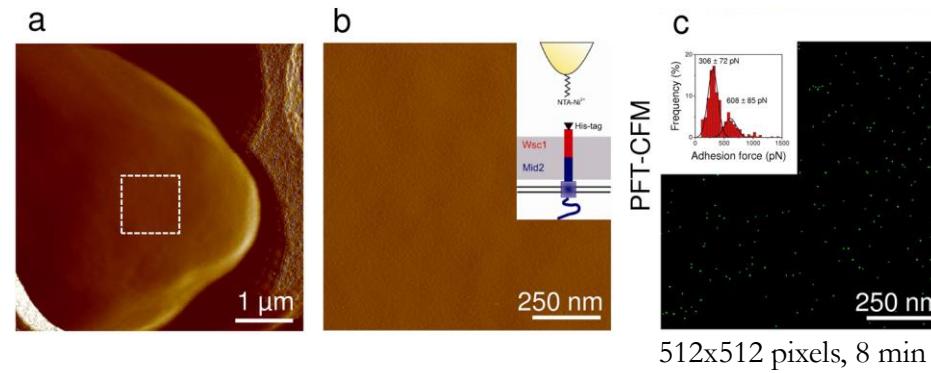
NTA-Ni²⁺ tips to detect His tagged proteins on live cells

- Revealing the spring behavior and clustering of Wsc1 in the yeast cell wall



Dupres *et al.* Nat. Chem. Biol. 2009

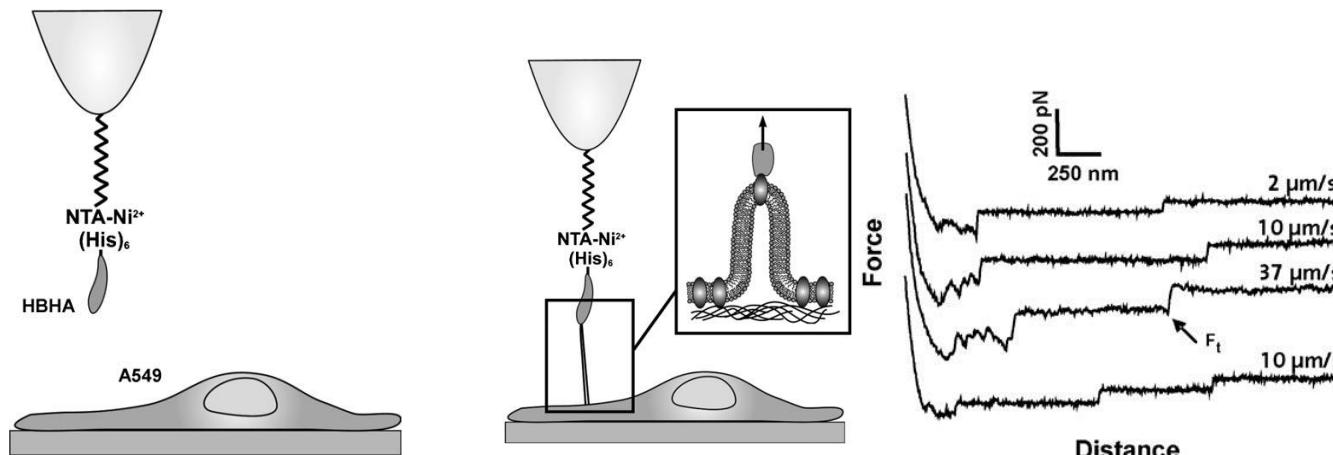
- High resolution detection of proteins using PFT-QNM



Alsteens *et al.* Langmuir 2012

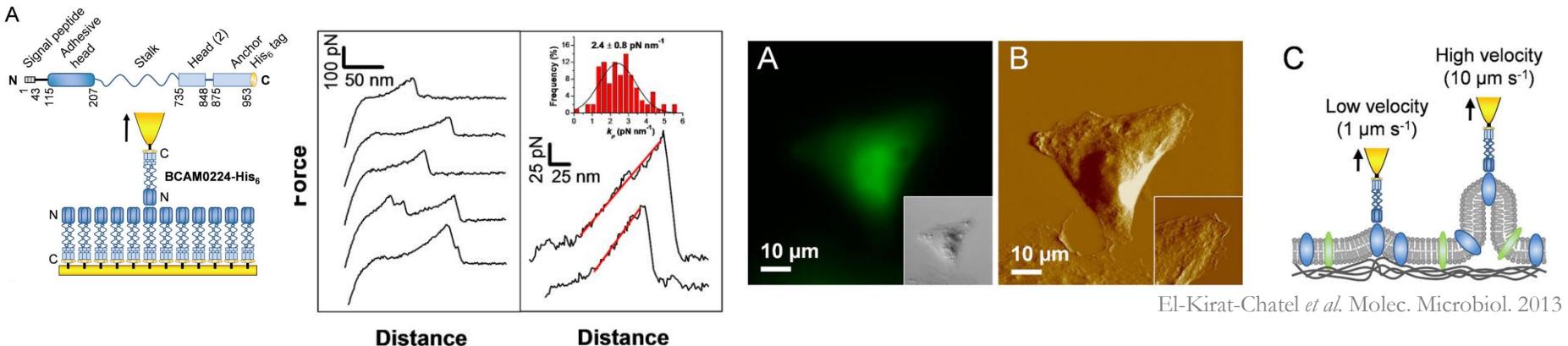
NTA-Ni²⁺ tips to graft oriented proteins and probe their interaction

- HBHA Mycobacterial adhesin recognize HSPG receptors on pneumocytes



Duprè *et al.* ChemPhysChem 2009

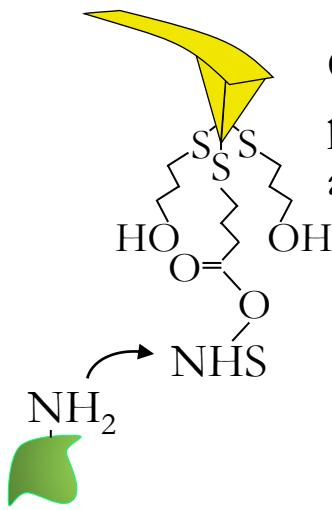
- BCAM *Burkholderia* TAA adhesin is involved in homotypic interaction and host recognition



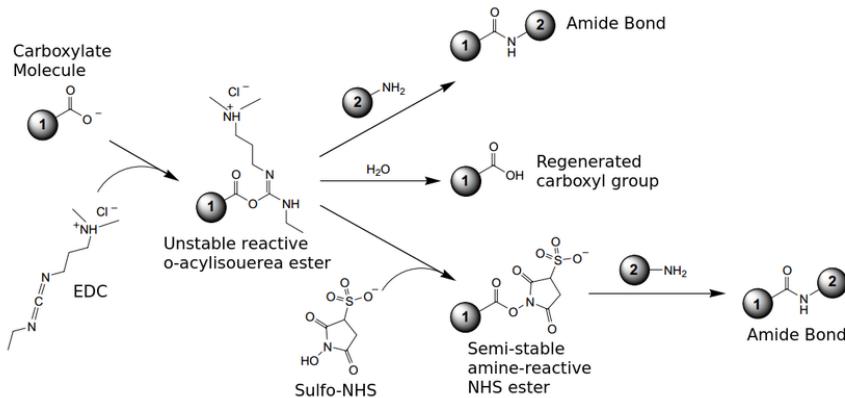
El-Kirat-Chatel *et al.* Molec. Microbiol. 2013

Limit: Functionalization reliable only if interaction force < 300 pN

Gold coated tip to graft
thiol-terminated molecules



Covalent grafting of
protein with primary
amines



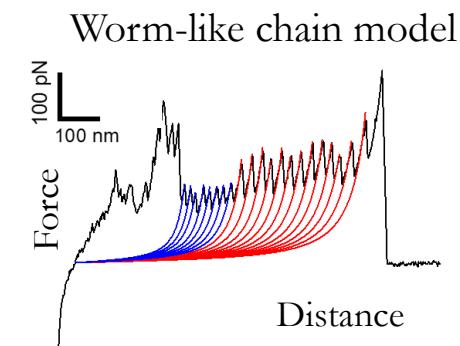
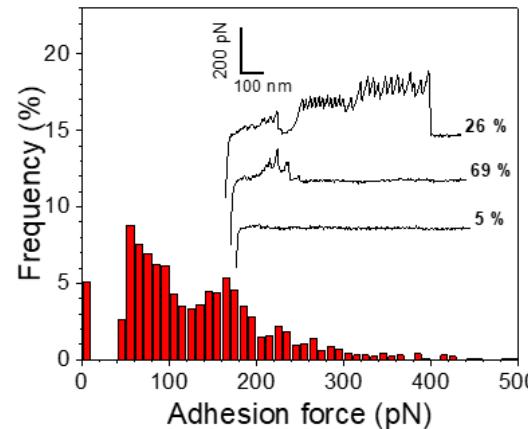
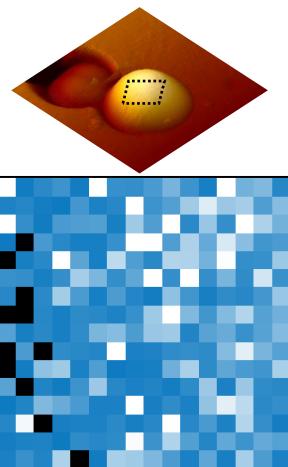
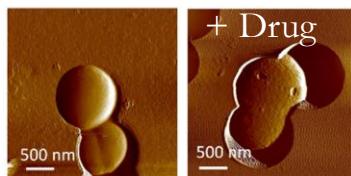
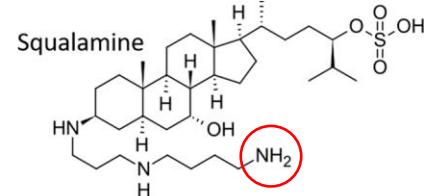
Hughes adapted from ThermoScientific

Requires highly pure proteins for reproducible results

Protocol steps:

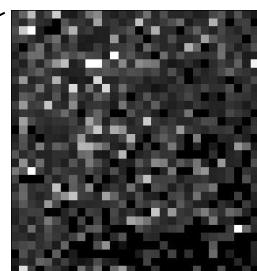
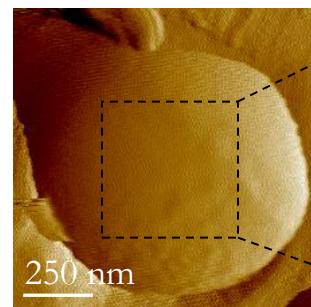
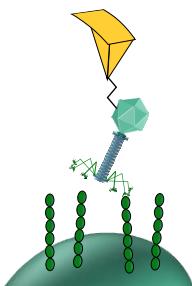
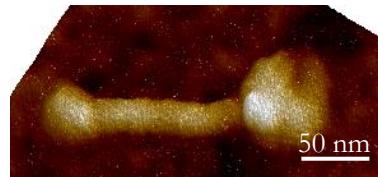
- AFM tip cleaning (UV/O_3 ; piranha...)
- Rinsing with ethanol + N_2 drying
- Immersion 4-12h in 0.1 mM 1-dodecanethiol and 0.9 mM SH-undecanol (=spacers)
- Rinsing with ethanol + N_2 drying
- Immerse 30 min in NHS-EDC (10mg/mL-25mg/mL in water)
- Rinse gently with pure water (avoid drying)
- Add 100-200 μ L of purified proteins (200 μ g/mL) 1-2h before gentle rinsing in buffer.

➤ Revealing nanoscale adhesion of antimicrobial molecules

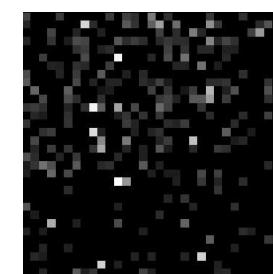


El-Kirat-Chatel *et al.* Col. Surf. B 2023

➤ Measuring phage-bacteria interaction



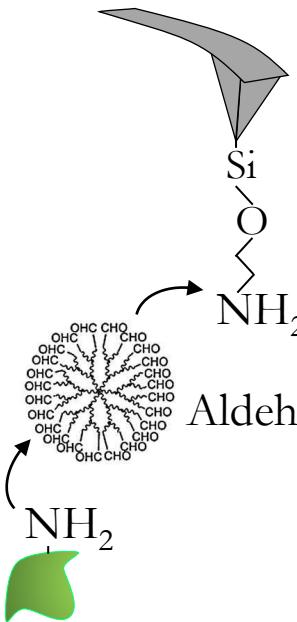
+ GlcNAc



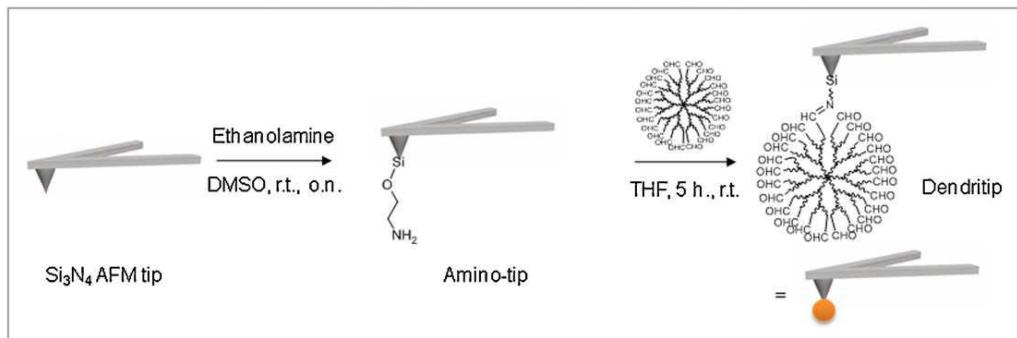
Arbez *et al.* Nano Res. 2022

Limit: Low molecular flexibility

Si_3N_4 tips decorated with amino groups



Covalent grafting of protein with primary amines



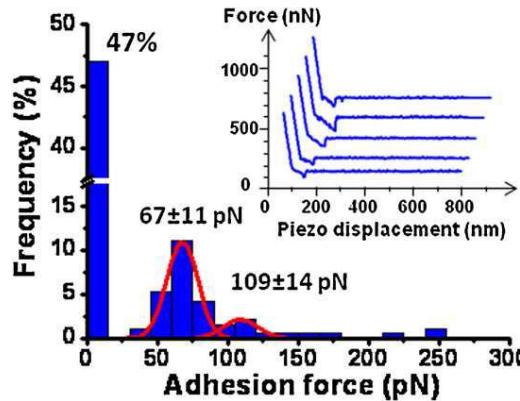
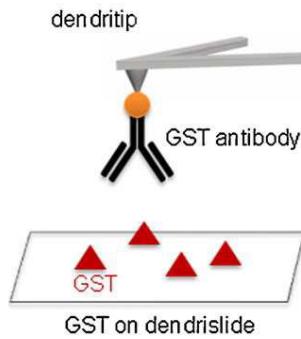
Jauvert et al. Sens. Actuators B Chem. 2012

Requires highly pure proteins or mAb

Protocol steps:

- AFM tip cleaning (Dichloromethane, acetone, ethanol, UV/ O_3)
- Immerse o/n in 0,5 mg/mL ethanalamine in DMSO+molecular sieve beads then rinse with DMSO and ethanol and dry with N_2
- Immerse 5h tetrahydrofuran containing dendrimer at 58 μM and rinse with DMSO and ethanol and dry with N_2
- Rinsing with ethanol + N_2 drying (possible to store in dissecator)
- Immerse 1h in 0,5mg/mL of proteins + 20 mM NaCNBH₃
- Rinse and store in buffer

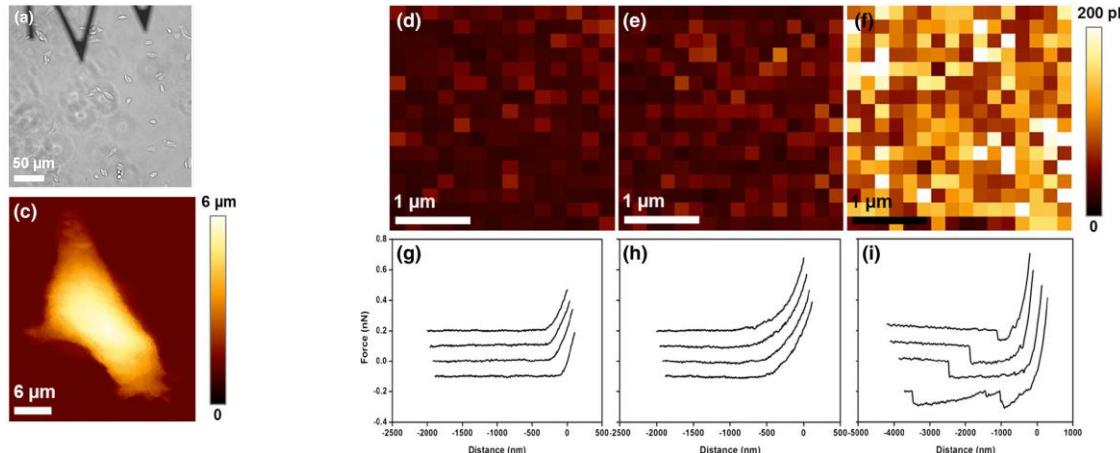
- Probing the interaction between glutathione-S-transferase (GST) and anti-GST mAb



→ Proof of concept

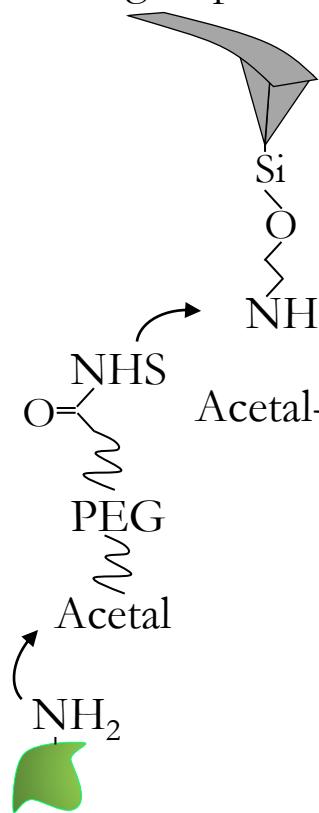
Jauvert *et al.* Sens. Actuators B Chem. 2012

- Detecting HA-tagged β 2-adrenergic receptors on CHO cells

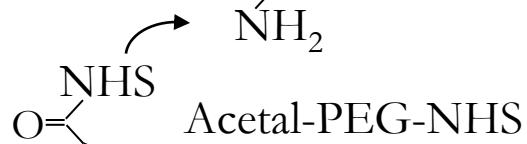


Flexibility, low unspecific binding BUT non oriented and NaCNBH_3 needed.

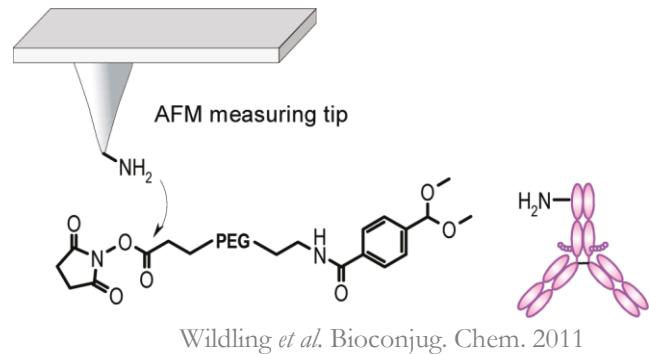
Si_3N_4 tips decorated with amino groups



Covalent grafting of protein with primary amines



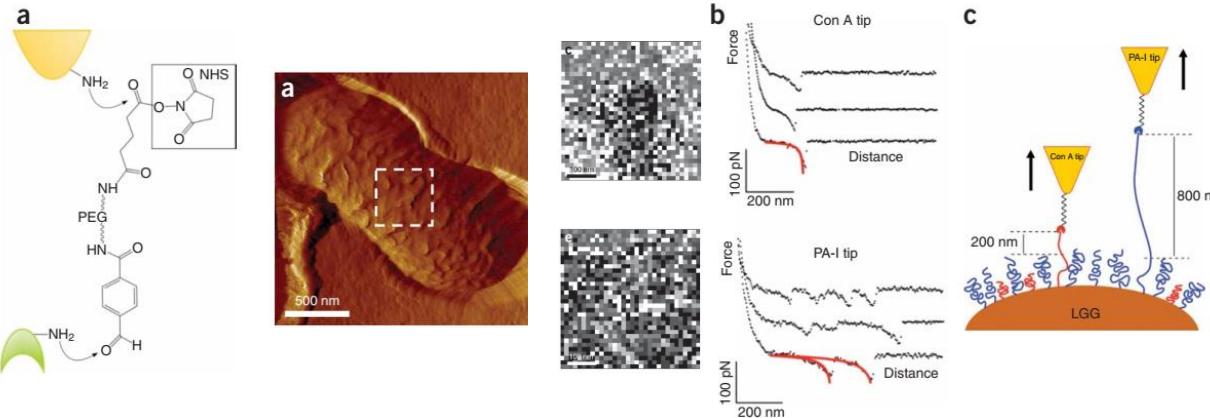
Requires highly pure proteins or mAb



Protocol steps:

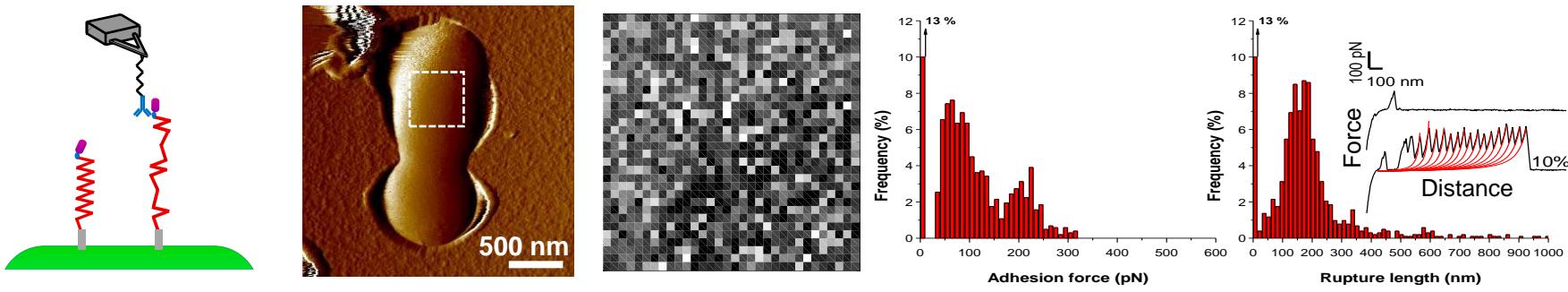
- AFM tip cleaning (Piranha?, UV/ O_3 , 3x chloroform)
- Immerse o/n in 0,5 mg/mL ethanolamine in DMSO+molecular sieve beads then rinse with DMSO and ethanol and dry with N_2
- Immerse 2h in 0,5mL chloroform + 1 mg Acetal-PEG-NHS
- Rinse with chloroform and dry with N_2 (possible to store in dissecator)
- Immerse 10 min in 1 % citric acid and rinse with water and dry with N_2
- Place cantilevers on parafilm
- Add ~100 μL of protein (~2 μM) + 2 μL of NaCNBH3 for 1h
- Add 5 μL of ethanolamine (1 M, pH 8.0) for 10 min
- Wash with buffer and store at 4°C

- Probing polysaccharides on bacteria



Francius *et al.* Nat. Protocol 2009

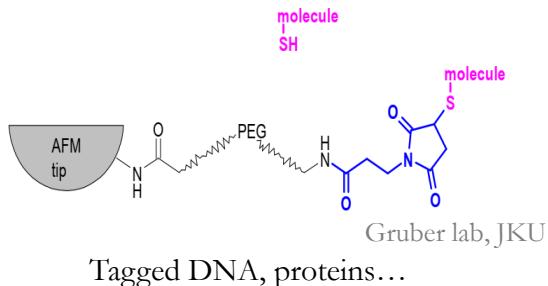
- Molecular biophysics of single bacterial adhesins



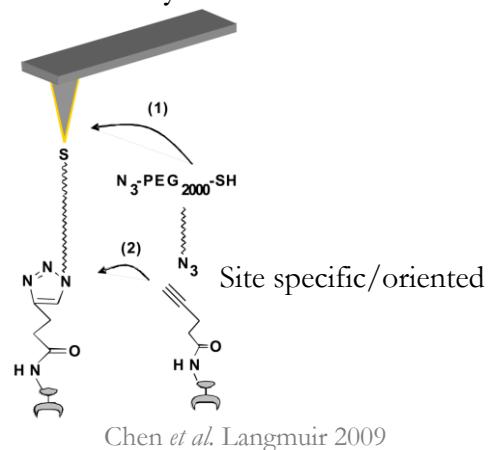
El-Kirat-Chatel *et al.* ACS Chem. Biol. 2013

Flexibility, low unspecific binding, specific FD signature BUT non oriented and NaCNBH₃ needed.

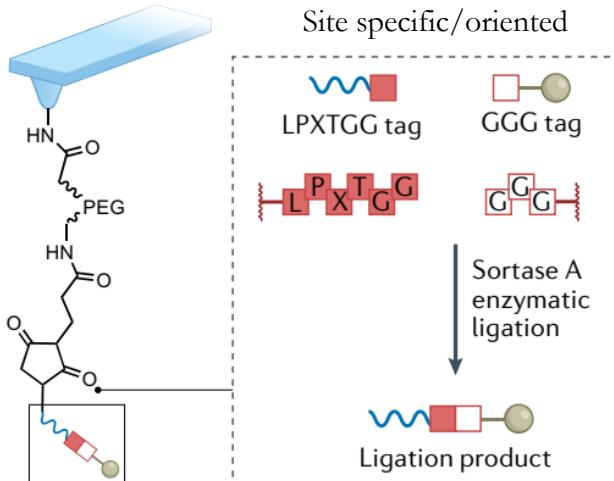
➤ Maleimide to graft –SH molecules



➤ Click chemistry



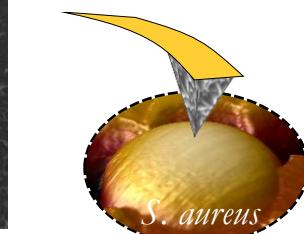
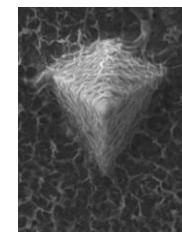
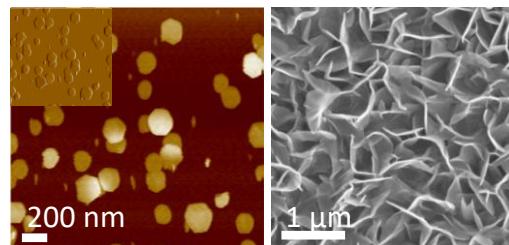
➤ LPXTGG/GGG/SortaseA from *S. aureus*



Theile *et al.* Nat. Protoc. 2013

Viljoen *et al.* Nat. Rev. Methods Primers 2021

➤ Particles: Layered double hydroxides

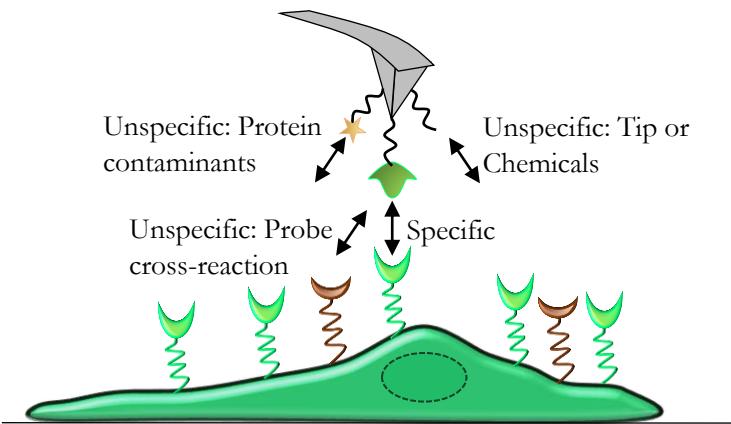


Probing antimicrobial LDH vs cells

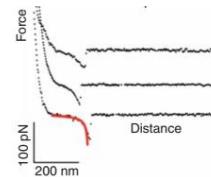
Awassa *et al.* Nanoscale 2022

➤ What are the control experiments?

Determining if the measured interaction is specific



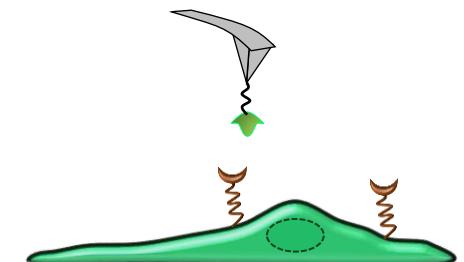
- Use highly pure protein preparation and avoid cross-reactive compounds
- Block unlinked linkers and add unreactive spacers
- Use poorly adhesive linkers (PEG...)
- Use linkers with specific extension signature (PEG → WLC)



Control experiments

Negative control sample (mutants):

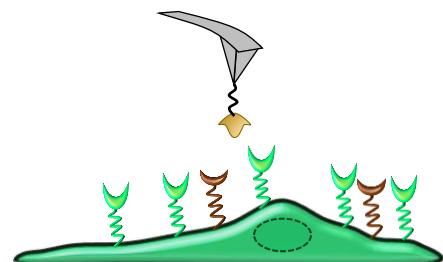
No adhesion



- No tip/chemical interaction
- No probe cross-reaction
- Reactive protein contaminant?
- Specificity

Aberrant probe:

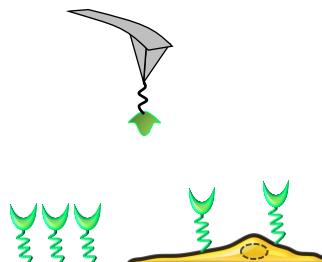
No adhesion



- No tip/chemical interaction
- No sample adhesion

Model surface/cell:

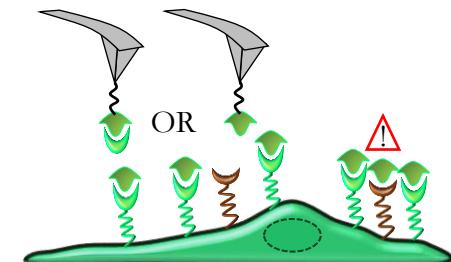
Adhesion



- Specificity
- FD shape

Blocking with free molecules:

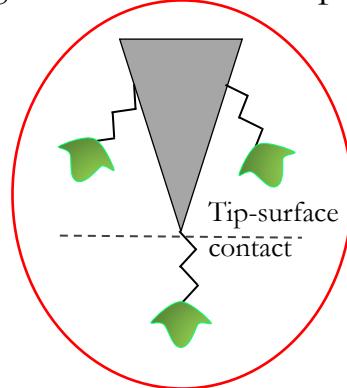
No adhesion



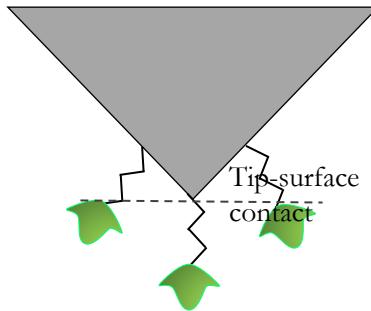
- No tip/chemical interaction
- Specificity (only with receptor injection, not with probe)

➤ How to determine if single-molecule interaction are probed?

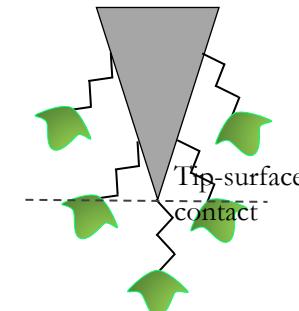
- Single-molecule force spectroscopy relies on the grafting density of molecules and the tip radius/sharpness



Only 1 molecule in contact with the surface

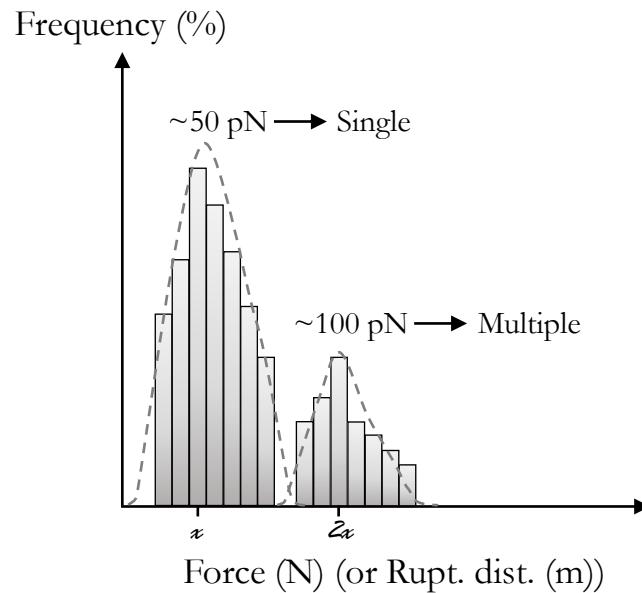
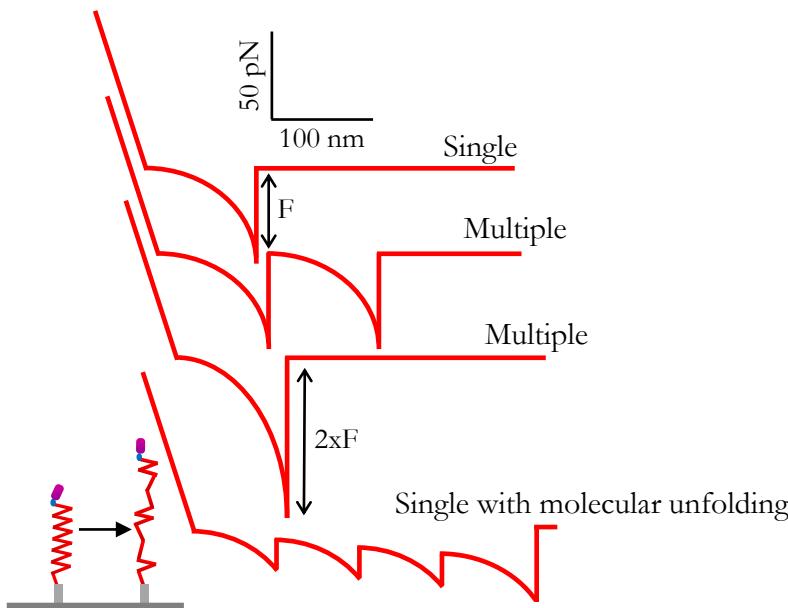


Tip too large



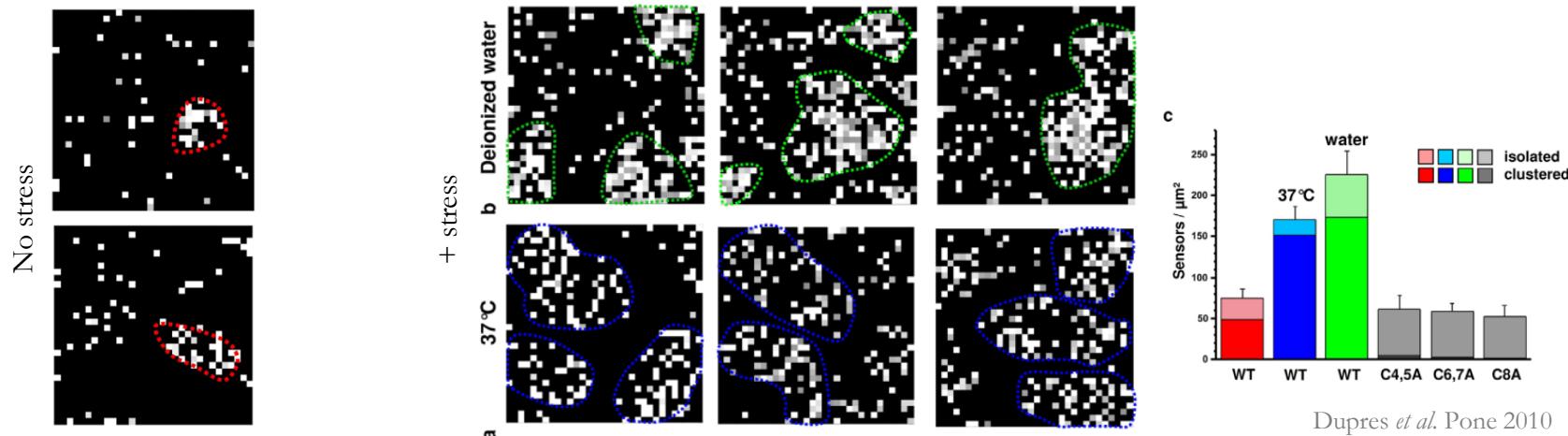
Density too high

- The force-distance curve signatures and the distribution of forces



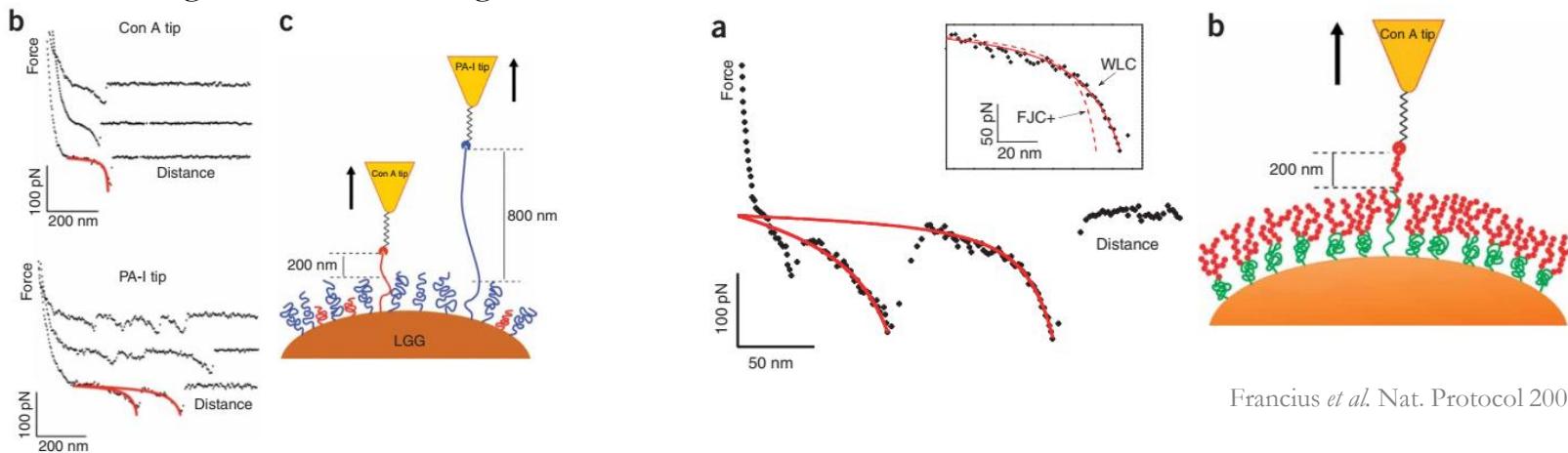
- What can be extracted from single-molecule force spectroscopy data?

- . Mapping: distribution, dynamics



Dupres *et al.* Pone 2010

- . Nature of the ligands: curve fitting



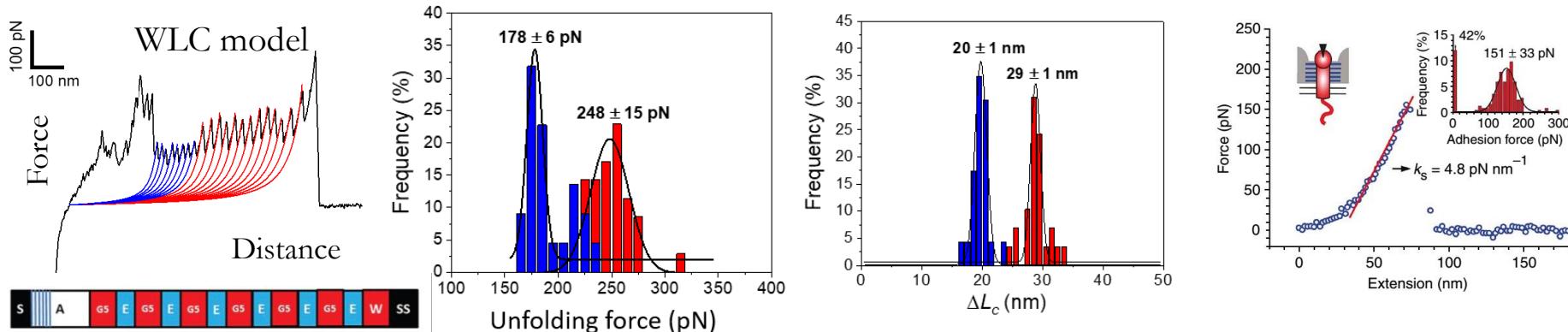
Francius *et al.* Nat. Protocol 2009

FJC and FJC⁺ for polysaccharides extension
(ideal chain with flexible joints)

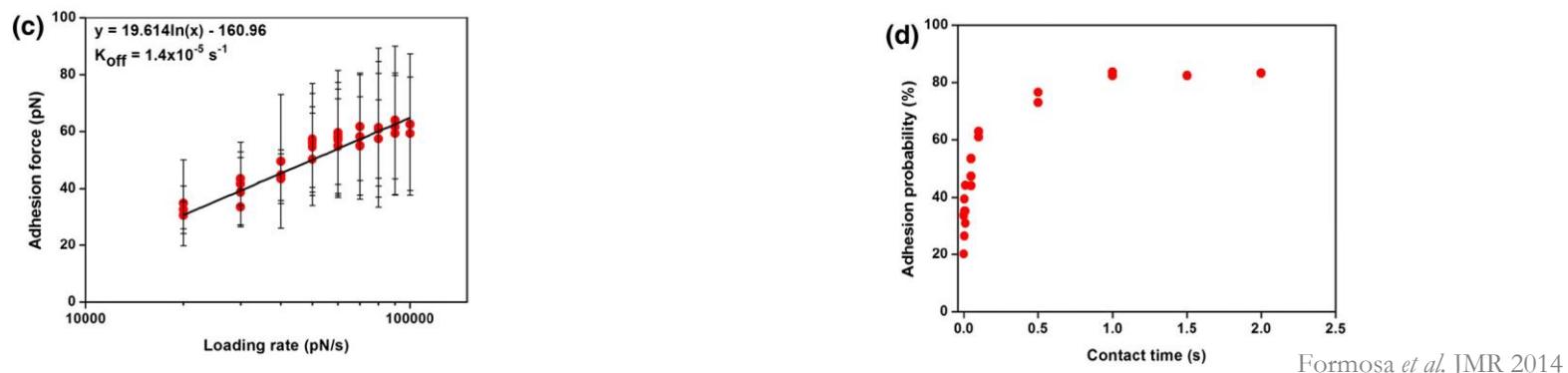
WLC and WLC⁺ for proteins and PEG extension
(continuously flexible chain irregularly curved)

➤ What can be extracted from single-molecule force spectroscopy data?

. Molecular mechanics: Unfolded domains? Adhesive domains? Nanosprings (linear fits)?



. Dynamic force spectroscopy (Bell-Evans; Dudko; Friddle *etc* models)

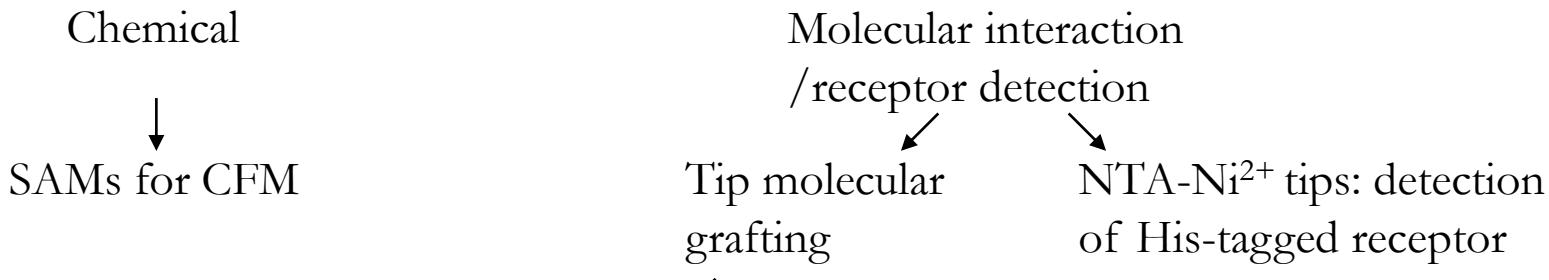


Increased loading rate by increasing pulling velocity
the dissociation constant (k_{off})

Increased contact time to determine the association
constant (k_{on})

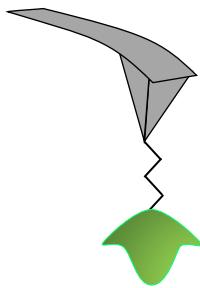
→ Affinity of the interaction k_d

1. What info I want to get?



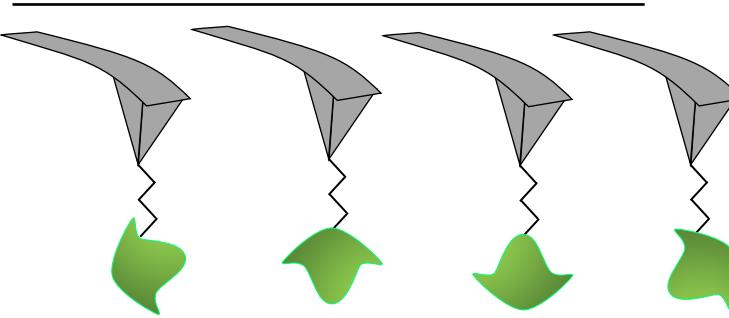
2. Orientation needed?

Yes



- Ni²⁺-NTA ($F < \sim 300 \text{ pN}$)
- LPXTGG/GGG/SortaseA from *S. aureus*
- Click chemistry

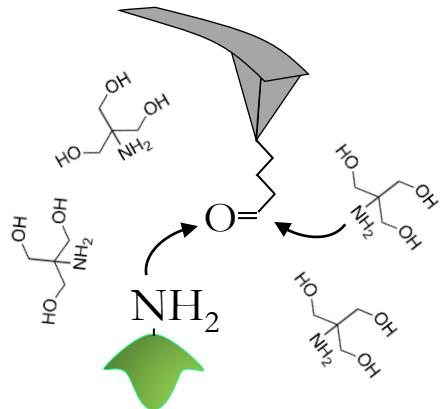
No



- All other covalent grafting through NH₂ groups (N-terminus and Lys side chains): NHS-EDC, PEG-linker, Dendritips... depending on the molecular flexibility and density required

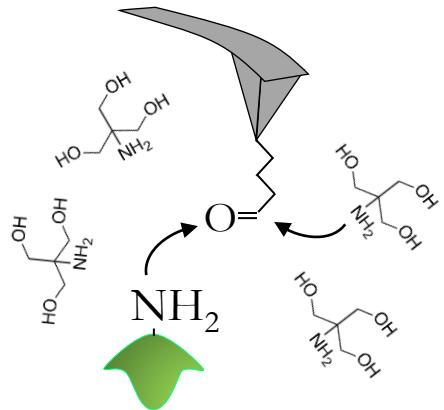
- + Maleimide linkers for -SH tagged molecules (Proteins, DNA). Oriented if only 1 -SH/molecule
- + Mineral chemistry for particles...
- + ...

3. Possible competition/cross-reactions?

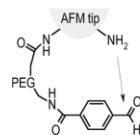


Do not use buffers containing
the coupling reactive group
(e.g. Tris for amine reactions)

3. Possible competition/cross-reactions?

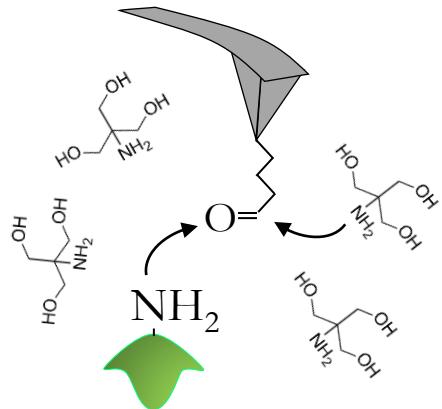


Do not use buffers containing
the coupling reactive group
(e.g. Tris for amine reactions)

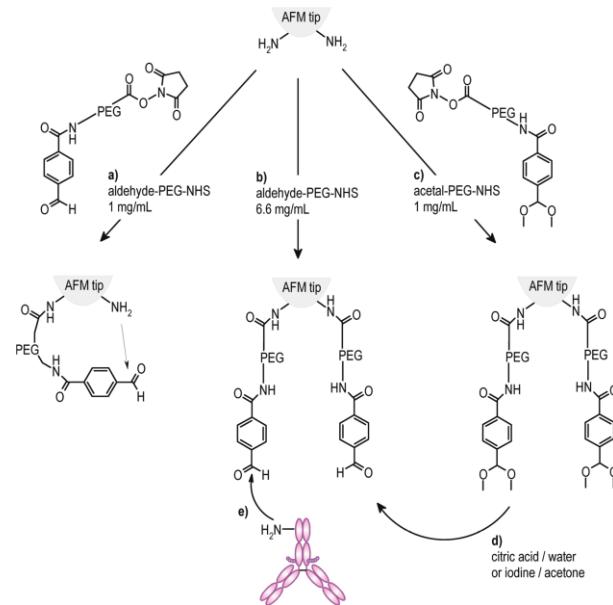


Blocking linker loops

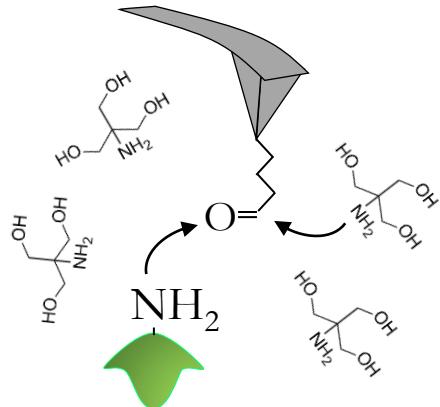
3. Possible competition/cross-reactions?



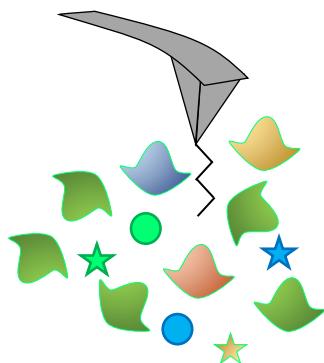
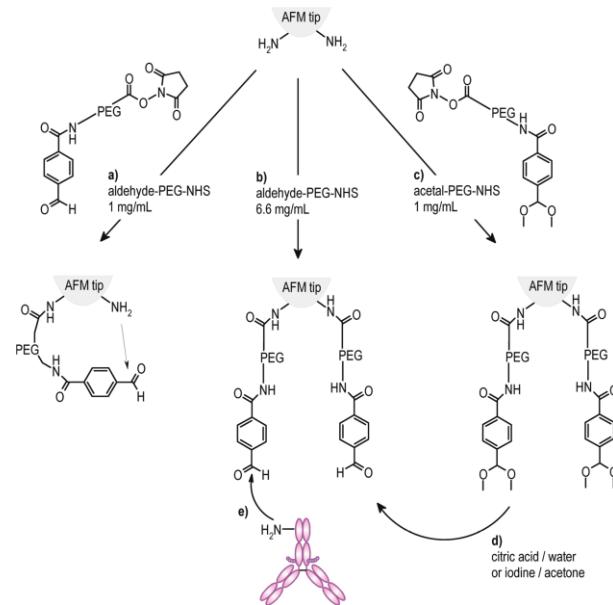
Do not use buffers containing
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3. Possible competition/cross-reactions?

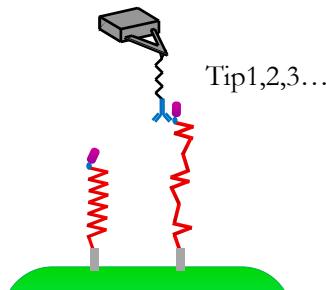


Do not use buffers containing
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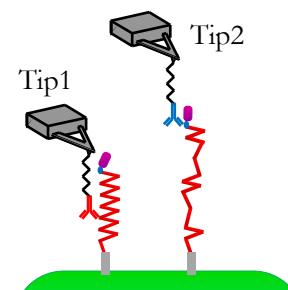


Always use highly pure
proteins for reproducible
results (purified proteins,
mAb...)

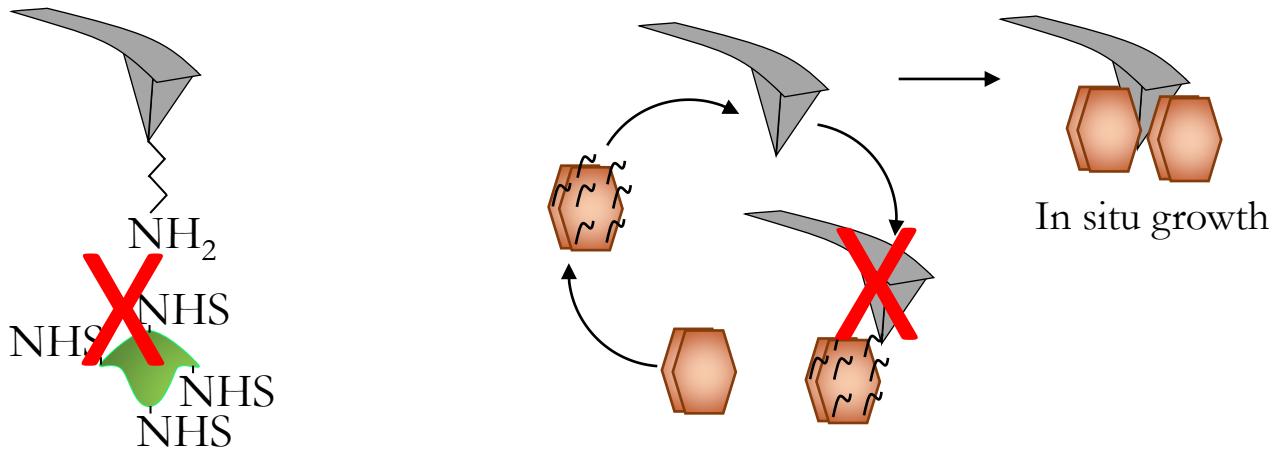
Monoclonal Ab:
Reproducible results
and force signatures



Polyclonal Ab:
Inconsistent results and
variable force signatures



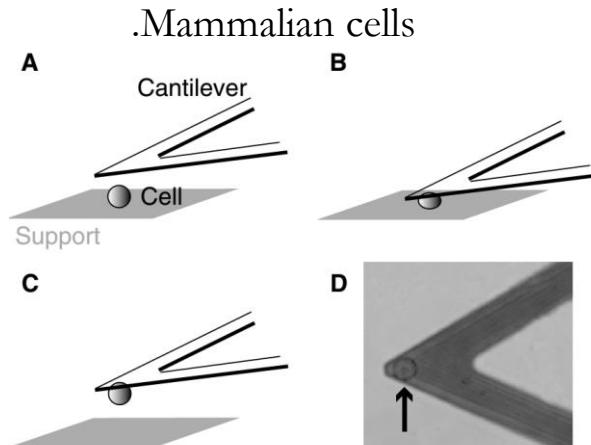
4. Probe modification/denaturation by the grafting?



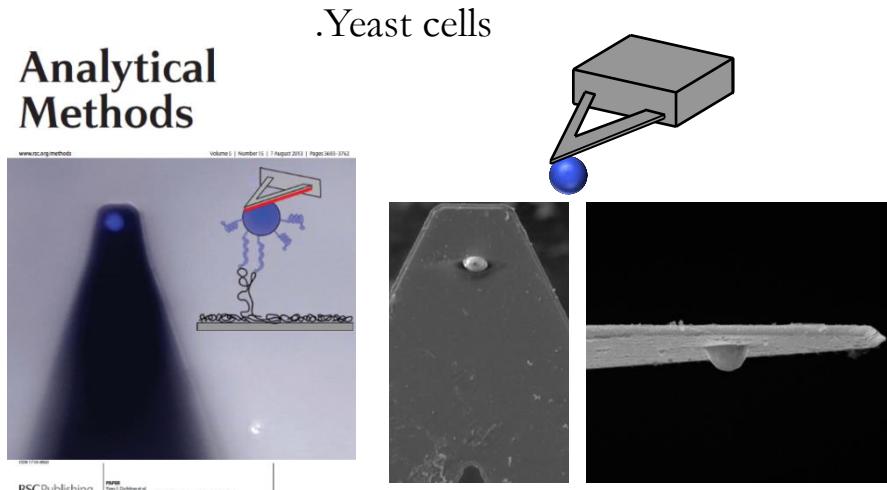
The protocol should modify the tip to add pure proteins. Avoid protein chemical modification/activation

Single-cell force spectroscopy: Attaching a single-cell to AFM cantilevers

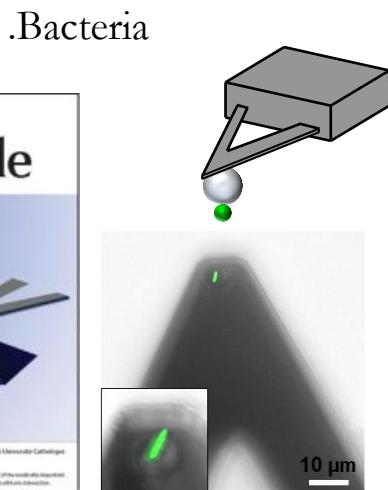
- Tipless cantilevers coated/functionalized with lectins or polydopamine OR FluidFM



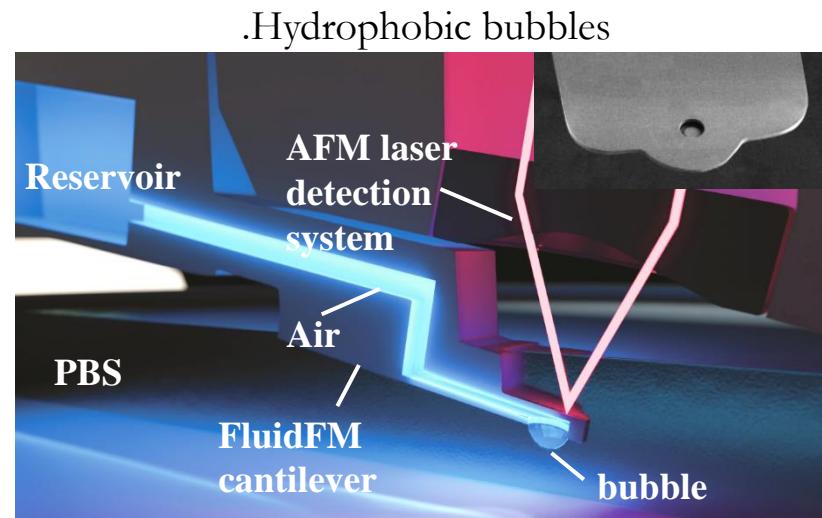
Helenius *et al.* J. Cell Sci. 2008



Alsteens *et al.* Analyt. Methods 2013



Beaussart *et al.* Nanoscale 2013



Demir *et al.* J. Coll. Interf. Sci. 2021

- For single-molecule force spectroscopy cantilevers should be sharp and soft enough to detect single-molecule interactions: $0,01 < k < 0,06 \text{ N/m}$; MLCT, SNL, NPG...
- Few suppliers for thiols, PEGs...: Sigma, Prochima, Broadpharm...
- Most protocols require delicate handling of probes for immersion in several small volume solutions poured in ultra clean glasswares.



- Few refs:
 - jku.at for pdfs on aminofunctionalization, acetal-PEG, maleimide-PEG...
 - Viljoen *et al.* Nat. Rev. Methods Primers 2021: Review on SMFS
(doi: 10.1038/s43586-021-00062-x)
 - Lostao *et al.* Internat. J. Biol. Macromolec. 2023: Review on SMFS
(doi: 10.1016/j.ijbiomac.2023.124089)
 - Bizzarri and Cannistraro Chem. Soc. Rev. 2010: Review on loading rate
(doi: 10.1039/b811426a)

Thank you for your attention



Enjoy your molecular fishing